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Alterations at the peptidyl transferase centre of the ribosome induced by the synergistic action of the streptogramins dalfopristin and quinupristin

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Abstract

Background

The bacterial ribosome is a primary target of several classes of antibiotics. Investigation of the structure of the ribosomal subunits in complex with different antibiotics can reveal the mode of inhibition of ribosomal protein synthesis. Analysis of the interactions between antibiotics and the ribosome permits investigation of the specific effect of modifications leading to antimicrobial resistances.

Streptogramins are unique among the ribosome-targeting antibiotics because they consist of two components, streptogramins A and B, which act synergistically. Each compound alone exhibits a weak bacteriostatic activity, whereas the combination can act bactericidal. The streptogramins A display a prolonged activity that even persists after removal of the drug. However, the mode of activity of the streptogramins has not yet been fully elucidated, despite a plethora of biochemical and structural data.

Results

The investigation of the crystal structure of the 50S ribosomal subunit from *Deinococcus radiodurans* in complex with the clinically relevant streptogramins quinupristin and dalfopristin reveals their unique inhibitory mechanism. Quinupristin, a streptogramin B compound, binds in the ribosomal exit tunnel in a similar manner and position as the macrolides, suggesting a similar inhibitory mechanism, namely blockage of the ribosomal tunnel. Dalfopristin, the corresponding streptogramin A compound, binds close to quinupristin directly within the peptidyl transferase centre affecting both A- and P-site occupation by tRNA molecules.
Conclusions

The crystal structure indicates that the synergistic effect derives from direct interaction between both compounds and shared contacts with a single nucleotide, A2062. Upon binding of the streptogramins, the peptidyl transferase centre undergoes a significant conformational transition, which leads to a stable, non-productive orientation of the universally conserved U2585. Mutations of this rRNA base are known to yield dominant lethal phenotypes. It seems, therefore, plausible to conclude that the conformational change within the peptidyl transferase centre is mainly responsible for the bactericidal activity of the streptogramins and the post-antibiotic inhibition of protein synthesis.
Background

Structural studies of complexes of both small and large ribosomal subunits with several clinically important antibiotics, for example, macrolides, lincosamides or chloramphenicol [1-3], have significantly advanced our understanding of the inhibitory action of these antimicrobial agents. However, the mechanism of the streptogramin class of antibiotics remains to be fully elucidated [4].

Streptogramins, which are produced by the genus *Streptomyces*, are divided into two types, A and B, both composed of macrocyclic lactone rings. Type B streptogramins (S\(_{B}\)) are cyclic hexadepsipeptides whereas type A (S\(_{A}\)) are highly modified cyclopeptides with multiple conjugated double bonds (Figure 1A). The antimicrobial activity of streptogramins has been well characterized (reviewed in [5]), revealing a unique, cooperative action. Alone, each compound exhibits a moderate bacteriostatic activity, but in combination the synergistic interplay between the compounds can produce a bactericidal effect [6, 7].

Type B streptogramins act on the 50S ribosomal subunit in a similar fashion as the macrolides and compete for the same binding site. The S\(_{B}\) do not affect the peptidyl transferase reaction, but inhibit elongation after a few cycles of peptide bond formation; by analogy with the macrolides, S\(_{B}\) are presumed to bind within the tunnel and block the path of the nascent polypeptide chain. Consistently, resistance to both classes of antibiotics arise through common mechanisms, for example, the resistance against macrolides, lincosamides and streptogramins B (MLS\(_{B}\)) results from methylation or mutation of A2058.
In contrast, $S_A$ act to prevent protein biosynthesis by blocking peptide bond formation. Type A streptogramins apparently do this by interfering with substrate binding at both acceptor and donor sites of the peptidyl transferase centre (PTC) [8], thereby inhibiting the peptidyl transferase reaction directly. Suppression of cell growth persists for a prolonged lag after removal of the drug [9, 10], presumably due to a stable perturbation of the conformation of the PTC induced by the binding of $S_A$ [11, 12].

The most recently approved streptogramin formulation is Synercid®, a 30:70 combination of dalfopristin ($S_A$) and quinupristin ($S_B$). The greatly enhanced solubility and bioavailability of Synercid®, and its excellent activity against Gram-positive as well as Gram-negative bacteria, has renewed interest in the medical use of the streptogramins.

To elucidate the unique cooperative effect of the streptogramin antibiotics, we investigated the structure of the 50S ribosomal subunit from *Deinococcus radiodurans* (D50S) in complex with both dalfopristin and quinupristin. The 3.4Å crystal structure allows the unambiguous localization of dalfopristin and quinupristin in the core region of the 50S ribosomal subunit, and enables for the first time a molecular understanding of the synergistic and prolonged inhibitory action of this clinically important class of antibiotics.
Results

Localization and interactions of quinupristin
Quinupristin is bound to 23S rRNA through an extensive network of hydrophobic interactions involving nucleotides of domain II, IV and V, hydrogen bonds between A2062 and C2586 (nucleotides are numbered according to the *E. coli* 23S rRNA sequence throughout the text), and the macrocyclic ring (Figure 1A, 2), in excellent agreement with biochemical and mutational data [5, 7, 12-30] (Figure 1B). Interestingly, the quinuclidinylthio-moiety (Figure 1A) occupies vacant space within the 50S subunit, suggesting that the addition of this moiety to the pristinamycin IA core enhances the bioavailability, but apparently not the binding properties, of quinupristin.

The binding site of quinupristin is located at the entrance to the ribosomal tunnel, and does not contact the active site of the 50S subunit (Figure 3A,B). This is in agreement with the observation that S_B does not affect peptide bond formation directly, but permits the ribosome to form a few peptide bonds until further extension of the nascent chain is prevented by blockage caused by binding of S_B. Quinupristin occupies the same space as the macrolides (Figure 3A), consistent with the strong competition observed between erythromycin and virginiamycin S (S_B) [25]. In fact, erythromycin can completely abolish S_B binding to the ribosome, illustrating the lower affinity of the S_R. Remarkably, in the *Haloarcula marismortui* 50S (H50S)-streptogramin complex no significant binding of S_B has been observed [4]. However, mutations of C2610U and especially A2058G, both of which are present in *H. marismortui*, have been found to confer resistance to S_B by reducing the binding affinity of the drug [31]; therefore, they provide a plausible explanation for the absence of S_B from the H50S complex.
Localization and interactions of dalfopristin
Dalfopristin is located in a tight pocket within the PTC, bound by a network of hydrophobic interactions involving the whole macrocyclic ring as well as the ethanethiol moiety (Figure 1, 2, 3B). The macrocyclic ring forms hydrogen bonds with G2505 and G2061, and the only hydroxyl contained in dalfopristin is hydrogen bonding to G2505. However, the most prevalent S_A resistance mechanism is based on the acetylation of this hydroxyl by virginiamycin acetyltransferase (VatD) [32]. Modification of the drug would, according to our structure, disrupt the hydrogen bond with G2505, and prohibit binding of acetylated S_A by steric hindrance. A large number of rRNA bases within the PTC have been shown to have altered reactivity in the presence of S_A [19, 21], among them G2058, A2059, A2439, G2505, U2506 and U2585, all of which directly contribute to the binding of dalfopristin (Figure 1B), demonstrating the agreement between structural and biochemical data.

The position of dalfopristin, which largely overlaps with that of chloramphenicol (Figure 3A), allows a clear interpretation of the inhibitory activity. Dalfopristin interferes with positioning of A- and P-site substrates (Figure 3A), although the CCA-end of tRNA might be flexible enough to bind unproductively even in the presence of S_A. But, once the P-site is occupied, binding of dalfopristin to the ribosome will be suppressed, thus explaining why ribosomes actively engaged in protein synthesis are not susceptible to S_A (reviewed in [5]). The loss of flexibility of the CCA-end in the P-site concomitant with nascent chain extension [33] might enhance this effect, since the effectiveness of S_A decreases as the number of amino acids attached to the P-site tRNA increases [30].
Discussion

Synergism

Synergistic binding of quinupristin and dalfopristin is presumably facilitated by strong hydrophobic interactions existing between both streptogramins, which lead to a significant reduction of their solvent accessible surface. Additionally, both compounds share contacts with a single nucleotide, A2062, through hydrophobic interactions as well as hydrogen bonds. Biochemical data implicate A2062 as undergoing conformational changes induced by streptogramin binding [16, 20]. When ribosomes were incubated with $S_A$ and $S_B$, the base of A2062, which was protected by $S_B$ alone, became accessible to dimethyl sulphate, suggesting that the conformational alterations of A2062 were induced by $S_A$ [16].

This proposal is further supported by the crystal structure of H50S, which contains two alternating conformations of A2062 [4]. In the complex of H50S with virginiamycin M ($S_A$), A2062 also appears to have undergone a conformational change, being rotated by $\sim 90^\circ$ compared with the prevalent orientation found in the native structure of H50S. But, as Hansen et al. (2003) [4] concluded, the conformational change of A2062 does not explain the post-antibiotic effect of $S_A$, since tRNA or substrate analogues bound to 50S induce a similar conformational change. In the native structure of D50S [34], A2062 obtains an orientation similar to the one observed in the H50S streptogramin complex (Figure 4A), but is rotated by $\sim 15^\circ$ in the D50S streptogramin complex to prevent steric clashes with $S_B$ and enable simultaneous accommodation of both $S_A$ and $S_B$. In this orientation, A2062 stacks between quinupristin and the macrocyclic ring of dalfopristin, such that any modification will severely affect the binding of both compounds. Mutations of A2062 are hence among the very few 23S rRNA modifications giving rise to both $S_A$ and $S_B$.
resistance [28]. The movement also affects the local environment, such that A2058 and A2060 are slightly displaced compared with the native structure (Figure 4A), which might contribute to the suppression of MLSB resistance by the presence of SA [11].

The most remarkable conformational changes induced within the PTC upon binding of dalfopristin are the alterations in the vicinity of U2585 (Figure 4B-D). This nucleotide, which points towards the tunnel in the native structure, appears to be rotated by about 180° to point in the opposite direction when the streptogramins are bound. This enables U2585 to form hydrogen bonds with C2606 and G2588 (Figure 4D), which should lead to a fairly stable alteration of the PTC.

To gain insight into the stability of this specific rRNA conformation, we performed a simple simulation by modelling a number of intermediates connecting the native and the streptogramin induced conformation of the region around U2585. Assuming that the transition does not induce large-range perturbations of 23S rRNA, the tightness of the region allows a single trajectory, which requires only moderate movements of A2439 to permit the continuous rotation of U2585 by 180°. Calculating the energy of each intermediate conformation yields a qualitative picture of the energy profile, showing that the transition requires the passage through an energy barrier (Figure 4E), to and from the native and streptogramin bound structures, which have essentially the same energy state. Binding of dalfopristin is obviously sufficient to force the native conformation of U2585 through the energy barrier to the alternate conformation, and spontaneous reversal of this transition will be comparably slow. This is in agreement with the observation that recovery of ribosomal activity after removal of SA is
followed by a prolonged lag [9, 10], suggesting that the post-antibiotic effect of $S_A$
can be attributed exclusively to the conformational change in U2585. Preliminary data
of a complex of D50S with the $S_A$ mikamycin A alone showed the same reorientation
of U2585 (data not shown), which confirms that the conformational change is neither
unique for dalfopristin nor dependent on the presence of $S_B$.

Although inversion of the orientation of U2585 appears to be most crucial for the
activity of $S_A$, the structure of the H50S-streptogramin complex [4] shows no
significant conformational change in U2585. This correlates with the peculiar
differences observed in the footprinting pattern of streptogramins on eubacterial and
archaeal ribosomes [21], and suggests that these antibiotics might exert a different
inhibitory mechanism between archea and eubacteria.

**Implications on peptide bond formation**
The universally conserved nucleotide U2585 has been implicated in the positioning of
P-site substrates by various biochemical experiments, and mutations of U2585 have
been shown to yield a lethal phenotype [35-39]. The recent structures of A- and P-site
substrates in complex with D50S [40] or H50S [41] placed U2585 in the direct
vicinity of the 3’ adenosine of the substrates and its attached amino acid (or mimics
thereof). The proposed mechanism for peptide bond formation, based on the local
two-fold symmetry inside the PTC, also suggests a direct interaction of the terminal
adenosine of P-site tRNA with U2585 and assigns a pivotal role for the rotary motion
during peptide bond formation to this nucleotide [42]. The correct conformation of
U2585 is hence essential for proper positioning of the substrate in the P-site [35-39],
which is a prerequisite for the formation of a functional 70S initiation complex. The
streptogramin induced conformation of U2585 should, therefore, severely affect the proper positioning of the 3' end of the P-site-bound tRNA, in agreement with biochemical data [16]. Conversely, interactions between U2585 and the P-site substrate might well stabilize the native conformation and hence suppress the conformational change required for Sₐ binding, contributing to the poor inhibition by Sₐ of ribosomes engaged in protein synthesis.

**Conclusions**

The structural studies of the 50S ribosomal subunit from *Deinococcus radiodurans* in complex with the streptogramins dalfopristin and quinupristin provides a first glance at the synergistic inhibition of protein synthesis. The bactericidal activity of the streptogramins can at least partially be attributed to the induction of the conformational change of the universally conserved nucleotide U2585. The altered conformation of U2585 has been shown to be stabilized by hydrogen bonds, leading to a rather stable distortion of the PTC. Therefore, it is likely that the conformational change is also responsible for the prolonged activity of Sₐ, which can persist for an extended period after removal of the drug. The synergistic inhibition appears to be a direct consequence of interactions between the two streptogramin components, but the fixation of A2062 in an orientation permitting simultaneous binding of both compounds contributes significantly to the synergistic activity.

**Methods**

**Base and amino acid numbering**

Nucleotides named (ACGU)1234 are numbered according to the 23S rRNA sequence from *E.coli* to permit a direct comparison with biochemical data. Translation tables converting *D. radiodurans* to *E. coli* to *H. marismortui* are available from the authors.
or at http://www.riboworld.com/nuctrans/. 23S rRNA sequence alignments were based on the 2D-structure diagrams obtained from Cannone et al. (2002) [43].

**Crystallization**

Crystals of the 50S ribosomal subunit were obtained as previously described by Harms et al. (2001) [34]. Co-crystallization was carried out in the presence of four-fold excesses of dalfopristin and a 10-fold excess of quinupristin.

**X-ray diffraction**

Data were collected at 85 K from shock-frozen crystals with synchrotron radiation beam at ID19 at Argonne Photon Source/Argonne National Laboratory (APS/ANL) and ID14/2, ID14/4 at the European Synchrotron Radiation Facility/European Molecular Biology Laboratory (ESRF/EMBL). Data were recorded on ADSC-Quantum 4 or APS-CCD detectors and processed with HKL2000 [44] and the CCP4 suite [45]. See Table 1 for data statistics.

**Localization and refinement**

The native structure of the 50S subunit was refined against the structure factor amplitudes of the 50S antibiotic complexes, using rigid body refinement as implemented in CNS [46]. For the calculation of the free R-factor, 5% of the data were omitted during refinement. The positions of the antibiotics were readily determined from sigmaA weighted difference maps (Figure 2C). The quality of the difference maps revealed unambiguously the position and orientation of the ligands. Since no structural model of quinupristin was available, virginiamycin S was used as
an initial model; the quinuclidinylthio-moiety was added manually and subsequently
minimized. The agreement between the initial model and the density was sufficient to
assume that the conformation of the macrocyclic ring of quinupristin and
virginiamycin S were identical. The placement of the quinuclidinylthio-moiety is
unambiguous; however, the density for this moiety is considerably weaker (invisible
contouring higher than 2.0σ) than the density of the core of the quinupristin structure
(still visible beyond 3.5σ) (Figure 2C). Further refinement was carried out using CNS
[46]. See Table 1 for refinement statistics.

Coordinates and figures
Three-dimensional figures were produced with Ribbons [47]. The ribosome-ligand
interactions were originally determined with LIGPLOT [48], but for sake of clarity
represented in a sketched manner. Final coordinates have been deposited in the
Protein Data Bank [49] under accession number 1SM1.

Authors' contributions
JMH modelled the structure and produced the images. FS performed the
computational tasks and drafted the manuscript. PF provided the materials and
crystals used in this study. FS and HB collected and processed the x-ray data. All
authors read and approved the final manuscript.

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Figure legends

Figure 1. Interactions of streptogramins with 23S rRNA
(A) Chemical structure of quinupristin and dalfopristin. The hydrogen bonds towards 23S rRNA nucleotides are indicated. (B) Overview of the nucleotides involved in binding in comparison with those indicated by various biochemical and genetic experiments [5, 7, 12-30]. Both images contain numbering for E. coli (in green) and for D. radiodurans (in red). The sequence itself corresponds to 23S rRNA of D. radiodurans. All other images use numbering according to E. coli.

Figure 2. Structure of dalfopristin and quinupristin within the PTC
To facilitate visualization of the interactions of dalfopristin and quinupristin with 23S rRNA, rRNA bases not involved in binding have been omitted. (A) Local environment of dalfopristin (in orange). A2062 is highlighted in purple; nucleotides, which are interacting through hydrogen bonds with either dalfopristin or quinupristin, are shown in dark blue. (B) Local environment of quinupristin (in green). Colours are as in (A). (C) Stereo view of the electron density map of quinupristin (in green) and dalfopristin (in orange). Both compounds have been omitted during calculation of the sigmaA weighted difference map, which is contoured at 1.5σ. (D) Stereo representation of dalfopristin and quinupristin and their local environment. Colours as in (A).

Figure 3. Comparison of antibiotic binding sites
(A) To visualize the relative orientations of different classes of antibiotics and substrates compared with dalfopristin and quinupristin, several structures have been aligned and overlaid: clindamycin (PDB entry 1JZX), erythromycin (1JZY),
chloramphenicol (1K01) and CC-Puromycin molecules in A- (A-CCPuro) and P-site (P-CCPuro). CC-Puromycin coordinates were taken from Bashan et al. (2003) [40].

(B) Overview of the binding sites of quinupristin and dalfopristin within the 50S ribosomal subunit, in relation to the P-site tRNA and the ribosomal exit tunnel (highlighted in gold).

**Figure 4. Conformation of the PTC**

(A) Different orientations of A2062 and its local environment for native D50S (purple), for the complex with Synercid® (blue) and for the H50S streptogramin complex (yellow). (B) Comparison of the position of dalfopristin in D50S (orange) with the position of virginiamycin M in H50S (light green), and the corresponding folds of 23S rRNA in the vicinity of U2585 (in blue for D50S and in yellow for H50S). (C) Structure and electron density around U2585. For comparison, the orientation of U2585 in the native D50S structure is also shown. The electron density is a sigmaA weighted difference map omitting the whole peptidyl-transferase ring from the calculation. (D) Local structure around U2585 overlaid with the native structure; putative hydrogen bonds of U2585 in its new orientation are indicated. (E) Approximate energy profile derived from modelling intermediate conformations, units being arbitrary. $N$ indicates the energy of the native conformation and $S$ the one of the complex with Synercid®.
### Tables

Table 1. Crystal parameters and statistics of data collection and refinement

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A

Dalfopristin

Quinupristin

Hydrogen bonds

ACGxxxx = D. radiodurans
xxxx = E. coli

B

CGAU Hydrogen bonds

Dalfopristin hydrophobic interactions
Quinupristin hydrophobic interactions

Dalfopristin biochemical or genetic data
Quinupristin biochemical or genetic data

ACGUxxxx = D. radiodurans
xxxx = E. coli